

The influence of *CAST/RsaI* and *RYR1* genotypes and their interactions on selected meat quality parameters in three groups of four-breed fatteners with different meat content of carcass*

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The effect was analysed of *CAST* and *RYR1* genotypes and their interactions on selected LL muscle quality traits in 201 fatteners of three four-breed crossbred groups. *CAST/RsaI* genotype affected WHC, drip loss at 96 h *post mortem* and protein and water content of muscle. Among analysed meat quality traits affected by *CAST/RsaI* genotype, animals of *AA* genotype were characterized by most desirable values.

Significant interactions between *CAST/RsaI* and *RYR1* genotype indicate that quality of meat should be considered not only as a result of genotype effect at each *locus*, but also as their combined effect.

KEY WORDS: *CAST* / fatteners /gene polymorphism / meat quality / *RYR1*

Up to year 2000 only an interactive effects of major genes (*RYR1* and *RN*) were studied on body composition and meat quality traits of pigs [Le Roy *et al.* 2000, Przybylski *et al.* 2000]. Recently, investigations were carried out concerning identification

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and effect of candidate genes [Emnet *et al.* 2000, Malek *et al.* 2001] and their interactions [Koćwin-Podsiadła *et al.* 2003].

The mentioned genes affect the rate of meat tenderization and post-slaughter protein turnover which is also influenced by an activity of calpastatin (CAST), an endogenous inhibitor of calpain [Goll *et al.* 1998]. Ciobanu *et al.* [2002] reported an effect of polymorphism in *CAST* gene identified with *Hpy*188I and *Pvu*II restriction enzymes on drip loss suggesting a necessity of further studies in order to explain whether the revealed effects were caused by analysed *CAST* gene mutations alone, or due to linkage disequilibrium.

The aim of this study was to analyse the effect of polymorphism of calpastatin (*CAST*) and *RYR1* genes (identified by digestion of PCR products with restriction endonuclease *Hin*PI for *RYR1* gene and *Rsa*I for calpastatin gene) on selected meat quality traits taking into consideration the meat content of carcass. Moreover, an effect was analysed of interactions between variants of *RYR1* and *CAST/Rsa*I genotypes and meat content of carcass on investigated meat quality traits.

Material and methods

Investigations were carried out on 201 crossbred fatteners representing three groups of crosses: [(Polish Large White × Polish Landrace) × (Duroc × Pietrain)] – 77 animals, [(Landrace × Yorkshire) × (Duroc × Pietrain)] – 40 animals, and [(Polish Large White × Polish Landrace) × (Hampshire × Pietrain)] – 84 animals. The animals were kept under similar environmental conditions, fed balanced diets and slaughtered using electrical pre-slaughter stunning (INARCO system) at a commercial abattoir, 4-5 hours after transportation on the distance 300 km. Immediately after slaughter blood samples were collected in EDTA-coated tubes for subsequent DNA analysis for the *RYR1* and *CAST* genotype.

Mean warm carcass weight (n=201) of analysed animals was 78.40 ±0.54 kg. Carcasses belonged to three groups of meat content: I – ≤50.0, II – 50.1 to 55 and III – >55%. Mean meat content of carcass was 51.28 ±0.40% (dissection made according to the Polish Pig Testing Stations method). In each group the number of gilts and castrated males was similar.

The following meat quality indicators were determined. pH of *Longissimus lumborum* (LL) muscle on the processing line (directly in carcass, in the region of the last rib) 35 min *post mortem* (pH₃₅) and in water homogenate of muscle tissue at 45 min (pH₄₅). R₁ – coefficient of energetic changes expressed as IMP/ATP ratio 45 min *post mortem* according to Honikel and Fischer [1977]. At 24 h *post mortem* meat lightness (measured with MINOLTA CR310 Chroma Meter in CIE L*a*b* system), water holding capacity (WHC) according to Grau and Hamm [1952] with Pohja and Niniivaara [1957] modification, and losses of weight of meat in cooking process. Drip loss from LL muscle tissue at 48 and 96 h post-slaughter was determined according to Prange *et al.* [1977]. Moreover, protein, water and dry matter contents of LL muscle tissue were

determined. Meat lightness and pH_{24} were measured also in *Semimembranosus* (SM) muscle. At min 45 *post mortem* samples from LL muscle were collected into tubes with 0.5 M PCA for determination of glycogen [Dalrymple and Hamm 1973] and lactate [Bergmeyer 1974]. On this basis the glycolytic potential (GP) was calculated according to formula proposed by Monin and Sellier [1985].

The *RYRI* genotypes were established according to Fujii *et al.* [1991]. Polymorphism of *CAST* gene was identified with *RsaI* endonuclease according to Ernst *et al.* [1998].

Statistical evaluation of the results was performed using three-way non-orthogonal ANOVA. Statistical model comprised *RYRI* and *CAST* genotypes, group of meat content of carcass and their interactions:

$$Y_{ijkl} = \mu + a_i + b_j + c_k + ab_{ij} + ac_{ik} + bc_{jk} + abc_{ijk} + e_{ijkl}$$

where:

- Y_{ijkl} – meat quality trait;
- μ – overall mean;
- a_i – effect of *CAST* genotype ($i=1,2,3$);
- b_j – effect of *RYRI* genotype ($j=1,2$);
- c_k – effect of the meat content of carcass ($k=1,2,3$);
- ab_{ij} – effect of interaction between *CAST* and *RYRI* genotypes;
- ac_{ik} – effect of interaction between *CAST* genotype and meat content group;
- bc_{jk} – effect of interaction between *RYRI* genotype and meat content group;
- abc_{ijk} – effect of interaction between *CAST* and *RYRI* genotypes and meat content group.
- e_{ijkl} – random effect.

Detailed comparison of means was made using Tukey test (STATISTICA PL 5.1).

Results and discussion

Observed and expected frequency of genotypes and alleles at the *CAST/RsaI* locus show that analysed population of animals was at Hardy-Weinberg equilibrium (Tab. 1).

Table 2 presents means and their standard errors of analysed meat quality traits, across genotypes considered and significance of their mutual interactions.

Highly significant ($P \leq 0.01$) influence of *RYRI* genotype on lactate level, pH_{35} and pH_{45} and significant on R_1 value ($P \leq 0.05$) were observed. It should be stressed that

Table 1. The frequency of genotypes and alleles at the *CAST/RsaI* locus

<i>RYR1</i>	<i>CAST/RsaI</i> (n=201)		
	<i>AA</i>	<i>AB</i>	<i>BB</i>
CC (n=114)	20	55	39
CT (n=87)	26	38	23
Total	46	93	62

Frequency of <i>CAST/RsaI</i> alleles (%)	
<i>A</i>	<i>B</i>
46.02	53.98

	Frequency of <i>CAST/RsaI</i> genotypes (%)		
	<i>AA</i>	<i>AB</i>	<i>BB</i>
Observed	22.89	46.27	30.84
Expected*	21.18	49.68	29.14
P value	ns	ns	ns

*According to Hardy-Weinberg equilibrium.

ns – difference between observed and expected frequency of genotype not significant.

animals considered were of *CC* and *CT* genotype at the *RYR1* locus. Related to *RYR1* genotype appeared also protein and dry matter content of muscle. In analysed fatteners' population highly significant influence of *RYR1* gene polymorphism was noted on pH_{35} , pH_{45} , R_1 values that are basis of PSE meat classification. As expected, the most desirable values of above mentioned parameters were noted in stress-resistant (*CC*) group of fatteners (Tab. 2).

A significant influence ($P \leq 0.05$) of *CAST/RsaI* genotype was noted for WHC, drip loss from muscle tissue 96 h *post mortem* and protein and water content. Analysing meat quality traits affected by *CAST/RsaI* genotype shows that *AA* animals were characterized by lower WHC (by 0.90 cm²), lower drip loss at 96 h (by 2.50 pp), lower water content (by 0.56 pp) and also by about 0.5 pp higher protein content compared to fatteners of *BB* genotype at this locus.

Significant interaction ($P \leq 0.05$) between *RYR1* and *CAST/RsaI* loci was noted for pH_{45} of LL muscle (Tab. 2, Fig. 1) and for drip loss at 48 h (Tab. 2, Fig. 2). No significant interactions were found between *CAST/RsaI* and *RYR1* genotype and group of meat content of carcass (Tab.2).

Interaction between all analysed factors (*CAST/RsaI*, *RYR1* and group of meat content) was significant ($P \leq 0.01$) for pH_{35} (Tab. 2, Fig.3).

Lee *et al.* [1992] showed that calcium channel activity might be regulated through domain L of calpastatin. It is also known that phosphorylase responsible for glycogenolysis is a substrate for calpain [Lametsch *et al.* 2002]. Moreover, degradation of glycogen in muscles *post mortem* may run with diverse speed in dependency on activity

Table 1. A relationship between polymorphisms at R1 and C43 (RsaI) and relations to meat quality parameters in pigs

Treat	R1/RsaI analysis				C43/RsaI analysis				Interaction analysis				
	RR	RR	RR	RR	CC	CC	CC	CC	RR/CC	RR/CC	RR/CC	RR/CC	
Lumbar level	46.07	43.24	46.08	46.08	45.00 ^a	42.14 ^b	--	43.24	43.24	44.08	--	44	44
	33.23	33.43	33.34	--	33.31	34.11	--	33.63	33.33	33.34	--	34	34
Dorsum	46.18	44.21	46.08	46.08	46.08	44.11	--	44.21	44.11	44.21	--	44	44
	33.81	32.83	33.08	--	33.81	33.23	--	33.83	33.23	33.03	--	34	34
Dorsal/loin/pelvic level	46.12	44.34	46.24	46.24	46.09	43.10	--	44.34	44.24	44.24	--	44	44
	34.08	33.53	33.83	--	33.92	33.11	--	33.58	33.44	33.23	--	34	34
pH ₂₄	6.23	6.49	6.41	6.41	6.24	6.43	--	6.31	6.43	6.43	--	64	64
	48.24	48.11	48.11	--	48.02	48.11	--	48.11	48.02	48.02	--	64	64
pH ₄₈ LL	6.29	6.23	6.24	6.24	6.11	6.14	--	6.24	6.18	6.18	--	64	64
	48.24	48.11	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
pH ₄₈ LL	5.93	5.93	5.93	5.93	5.74	5.91	--	5.93	5.93	5.93	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
pH ₄₈ RL	5.23	5.23	5.24	5.24	5.23	5.23	--	5.24	5.23	5.23	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
E ₁	0.81	0.83	0.83	0.83	0.80	0.81	--	0.80	0.83	0.81	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
Sawtooth index	50.24	50.23	50.24	50.24	50.28	50.28	--	50.24	50.23	50.23	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
H ₁ (mm)	48.02	48.02	48.02	48.02	48.02	48.02	--	48.02	48.02	48.02	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
Sawtooth index	48.02	48.02	48.02	48.02	48.02	48.02	--	48.02	48.02	48.02	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
Sawtooth index	48.02	48.02	48.02	48.02	48.02	48.02	--	48.02	48.02	48.02	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
Sawtooth index	48.02	48.02	48.02	48.02	48.02	48.02	--	48.02	48.02	48.02	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
Sawtooth index	48.02	48.02	48.02	48.02	48.02	48.02	--	48.02	48.02	48.02	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64
Sawtooth index	48.02	48.02	48.02	48.02	48.02	48.02	--	48.02	48.02	48.02	--	64	64
	48.02	48.02	48.02	--	48.02	48.02	--	48.02	48.02	48.02	--	64	64

Table 1. Continued

trial	Control condition				PTD condition				Dissolved condition				Fundamental condition						
	dd	df	df	sqd.	cc	cc	cc	sqd.	%0%	10.00%	20%	sqd.	cc	cc	cc	sqd.	cc	cc	sqd.
Group mean of %s	6.07	3.27	9.26	-	5.18	3.84	9.41	-	5.17	6.79	9.41	-	5.17	6.79	9.41	-	5.17	6.79	9.41
[M]	(0.28)	(0.25)	(0.24)	-	(0.22)	(0.21)	(0.21)	-	(0.22)	(0.21)	(0.21)	-	(0.22)	(0.21)	(0.21)	-	(0.22)	(0.21)	(0.21)
Total probes	20.07	22.16	22.27	-	22.28	22.27	-	22.27	22.22	22.22	22.22	-	22.27	22.22	22.22	-	22.27	22.22	22.22
standard [M]	(0.22)	(0.24)	(0.22)	-	(0.22)	(0.22)	-	(0.22)	(0.22)	(0.22)	(0.22)	-	(0.22)	(0.22)	(0.22)	-	(0.22)	(0.22)	(0.22)
Word length	9.22	9.10	9.20	-	9.18	9.07	-	9.16	9.17	9.17	9.17	-	9.16	9.17	9.17	-	9.16	9.17	9.17
[M]	(0.25)	(0.25)	(0.25)	-	(0.21)	(0.21)	-	(0.21)	(0.21)	(0.21)	(0.21)	-	(0.21)	(0.21)	(0.21)	-	(0.21)	(0.21)	(0.21)
Big words	26.42	27.16	26.26	-	27.20	26.29	-	27.22	26.29	26.29	26.29	-	27.22	26.29	26.29	-	27.22	26.29	26.29
standard [M]	(0.18)	(0.21)	(0.22)	-	(0.21)	(0.21)	-	(0.21)	(0.21)	(0.21)	(0.21)	-	(0.21)	(0.21)	(0.21)	-	(0.21)	(0.21)	(0.21)

cc - non-significant, P > 0.05, **P < 0.01, ***P < 0.001
 df - within row, means bearing different superscripts differ significantly at a level $\alpha = 0.05$; sqd/sqd - PTDD1
 CCM - group of non-concrete nouns
 LL - long/short adjectives/adverbs
 SM - concrete/abstract nouns

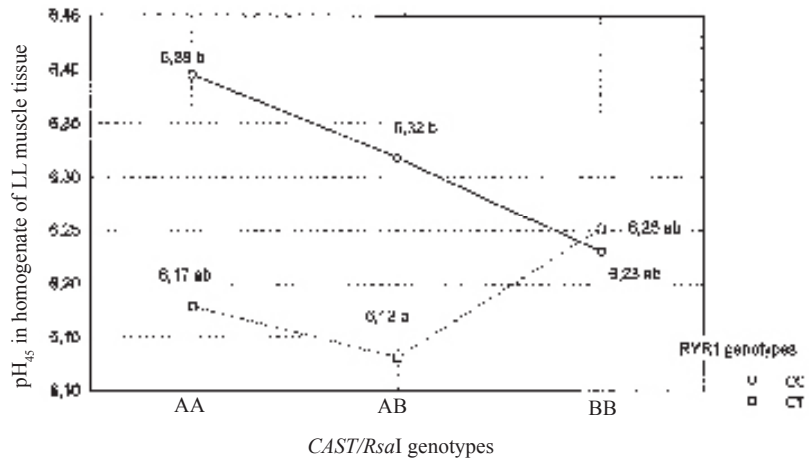


Fig. 1. Interactive effect of *CAST/RsaI* and *RYR1* genotypes for pH₄₅ in homogenate of LL muscle tissue.

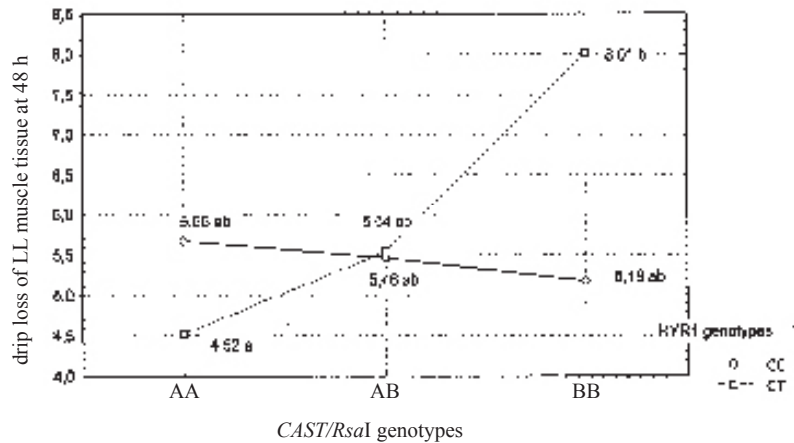


Fig. 2. Interactive effect of *CAST/RsaI* and *RYR1* genotypes for drip loss of LL muscle tissue at 48 h.

of these proteases regulated by calpastatin and it may explain the effect of *CAST/RsaI* × *RYR1* interaction on pH₄₅ value (Fig. 1).

Differentiation of drip loss from LL muscle tissue at 48 h was observed only among animals heterozygous (*CT*) at *RYR1* locus (Fig. 2). Drip loss from meat of *CT* animals being simultaneously *BB* homozygotes at *CAST/RsaI* locus was about 3.5 pp higher than that in group of the same *RYR1* genotype but *AA* homozygous at *CAST/RsaI* locus (4.52% and 8.01% respectively).

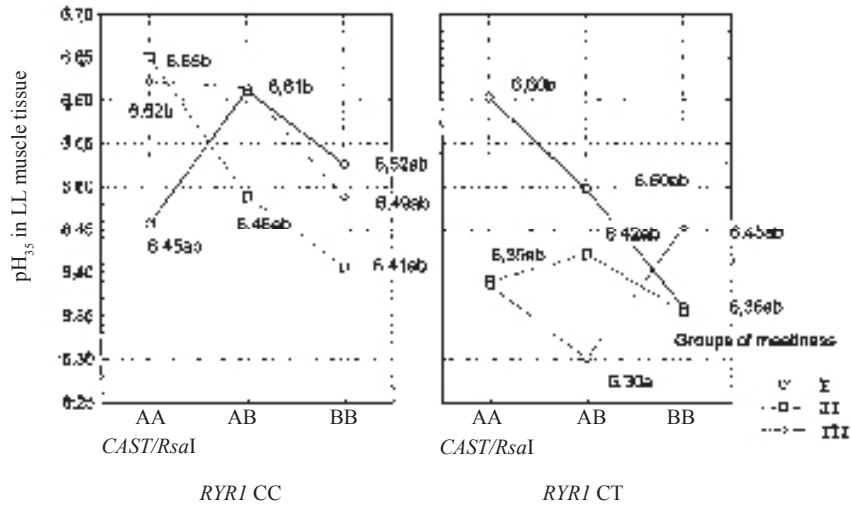


Fig. 3. Interactive effect of *CAST/RsaI* and *RYRI* genotypes with group of meatiness for pH_{35} in LL muscle tissue.

Obtained interaction between *CAST/RsaI* and *RYRI* genotypes and group of meat content of carcass indicate that presence of *T* allele of *RYRI* gene in heterozygous form may cause differentiation in pH_{35} of LL muscle among animals with meat content above 55%, being *AB* heterozygotes at *CAST/RsaI* locus (Fig. 3).

The polymorphisms of the *CAST* gene genotyped in this study were located in intron 6 and it is difficult to conclude on their effect on calpastatin level or activity. Nevertheless, Le Hir *et al.* [2003] showed the important role of introns in eukariotic genes on influencing gene expression by increasing transcriptional efficiency of numerous genes.

Significant interactions found between *CAST/RsaI* and *RYRI* genotypes indicate that quality of meat should be considered as not only a result of influence of genotype at each locus separately, but also as an effect of their interactions.

Summing up, the results presented suggest that relationships between polymorphism in *CAST* gene and meat quality of pigs should be further investigated.

REFERENCES

1. BERGMAYER H.U., 1974 – Methods of enzymatic analysis. Academic Press, New York.
2. CIOBANU D.C., LONERGAN S.M., BASTIAANSEN J.W.M., WOOLLARD J.R., MALEK M., HUFF-LONERGAN E.J., PLASTOW G.S., ROTHSCILD M.F., 2002 – Evidence for new alleles in calpastatin gene associated with meat quality traits in pigs. Proceedings of the 7th World Congress on Genetics Applied to Livestock Production. Montpellier (France), 19-23 August, Book of Abstracts, 19-23.
3. DALRYMPLE R.H., HAMM R., 1973 – A method for the extraction of glycogen and metabolites from a single muscle sample. *Journal of Food Technology* 8, 439-444.

4. EMNETT R., MOELLER S., IRVIN K., ROTHSCCHILD M., PLASTOW G., GOODWIN R., 2000 – An investigation into the genetic controls of pork quality. Proceedings of the 25th Anniversary of the NSIF Conference and Annual Meeting, Nashville, Tennessee, (USA), 7-8 December <http://mark.asci.ncsu.edu/nsif/00proc/emnett.htm>
5. ERNST C.W., ROBIC A., YERLE M., WANG L., ROTHSCCHILD M.F., 1998 – Mapping of calpastatin and three microsatellites to porcine chromosome 2q2.1-q2.4. *Animal Genetics* 29, 212-215.
6. FUJII J., OTSU K., ZORZATO F., DE LEON S., KHANNA S., WEILER V.K., O'BRIEN P.J., MACLENNAN D.H., 1991 – Identification of a mutation in porcine ryanodine receptor associated with malignant hyperthermia. *Science* 253, 448-451.
7. GOLL D.E., THOMPSON V.F., TAYLOR R.G., OUALI A., 1998 – The calpain system and skeletal muscle growth. *Canadian Journal of Animal Science* 78, 503-512.
8. GRAU R., HAMM R., 1952 – Eine einfache Methode zur Bestimmung der Wasserbindung in Fleisch. *Fleischwirtschaft* 4, 295-297.
9. HONIKEL K.O., FISCHER H., 1977 – A rapid method for the detection of PSE and DFD porcine muscles. *Journal of Food Science* 42, 1633-1636.
10. HONKAVAARA M., 1997 – Evaluation of the frequency of PSE pork in Finnish slaughter pigs and its technological effects. Proceedings of the 43rd International Congress of Meat Science and Technology, Auckland (New Zeland), 27 July-01 August, 296-297.
11. KOĆWIN-PODSIADŁA M., KURYŁ J., KRZĘCIO E., ZYBERT A., PRZYBYLSKI W., 2003 – The interaction between calpastatin and RYR1 genes for some pork quality traits. *Meat Science* 65, 731-735.
12. LAMETSCH R., ROEPSTORFF P., BENDIXEN E., 2002 – Identification of protein degradation during post-mortem storage of pig meat. *Journal of Agricultural and Food Biochemistry* 50, 5508-5512.
13. LEE W.J., MA H., TAKANO E., YANG H.Q., HATANAKA M., MAKI M., 1992 – Molecular diversity in amino-terminal domains of human calpastatin by exon skipping. *Journal of Biological Chemistry* 267, 8437-8442.
14. LE HIR H., NOTT A., MOORE M.J., 2003 – How introns influence and enhance eucaryotic gene expression. *Trends in Biochemical Sciences* 28 (4), 216-220.
15. LE ROY P., MORENO C., ELSEN J.M., CARITEZ J.C., BILLON Y., TALMANT A., VERNIN P., AMIQUES Y., SELLIER P., MONIN G., 2000 – Interactive effects of the HAL and RN major genes on carcass quality traits in pigs preliminary results. In: Quality of meat and fat in pigs as affected by genetics and nutrition (C. Wenk, J.A. Fernandez and M. Dupuis M., Eds). Wageningen Pers, Wageningen, 139-142.
16. MALEK M., DEKKERS J.C.M., LEE H.K., BAAS T.J., PRUSA K., HUFF-LONERGAN ROTHSCCHILD M.F., 2001 – A molecular genome scan analysis to identify chromosomal regions influencing economic traits in the pig. I. Growth and body composition. *Mammalian Genome* 12, 637-645.
17. MONIN G., SELLIER P., 1985 – Pork of low technological quality with a normal rate of muscle pH fall in the immediate post-mortem period: The case of Hampshire breed. *Meat Science* 13, 49-63.
18. POHJA N.S., NINIVAARA F.P., 1957 – Die Bestimmung der Wasserbindung des Fleisches mittels der Konstandrückmethods. *Fleischwirtschaft* 9, 193-195.
19. PRANGE H., JUGRRT L., SCHARNER E., 1977 – Untersuchungen zur Muskel fleischqualität beim Schwein. *Archiv für Experimentelle Veterinarmedizin*, Leipzig 31(2), 235-248.
20. PRZYBYLSKI W., KOĆWIN-PODSIADŁA M., KACZOREK S., KRZĘCIO E., MONIN G., 2000 – Interactive effects of the HAL and RN genes in crossbreeding. Proceedings of the 46th International Congress of Meat science and Technology, Buenos Aires (Argentina), 27 August-1 September, 2.I-P18, 88-89.

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Wpływ genotypów *CAST/RsaI* i *RYRI* oraz ich interakcji na wybrane parametry jakości wieprzowiny w trzech grupach czterorasowych mieszańców, zróżnicowanych zawartością mięsa w tuszy

Streszczenie

Celem badań była analiza wpływu genotypów *CAST/RsaI* i *RYRI* oraz ich interakcji na cechy jakości mięśnia *longissimus lumborum*. Badaniami objęto 201 tuczników trzech grup czterorasowych mieszańców. Genotyp *CAST/RsaI* istotnie wpływał na wodochłonność mięsa (WHC), naturalny wyciek w 96 godzinie *post mortem* oraz zawartość białka i wody w mięsie. Korzystniejszą wartość tych cech wykazywały zwierzęta o genotypie *AA*. Wykazany istotny wpływ interakcji genotypów *CAST/RsaI* i *RYRI* na wartość niektórych parametrów charakteryzujących jakość mięsa wskazuje, że są one kształtowane nie tylko przez genotyp względem każdego z analizowanych *loci*, ale również przez współdziałanie między tymi *loci*.